# Intrahepatic CD8+ T cells correlate with significant declines in HBV viral load and S antigen following a single vaccination with VRON-0200 in an AAV mouse model

M HASANPOURGHADI,<sup>1</sup> <u>A LUBER</u>,<sup>2</sup> S CURRIE,<sup>2</sup> X ZHOU,<sup>1</sup> and HCJ ERTL<sup>1</sup>

<sup>1</sup>The Wistar Institute, Philadelphia, PA, USA; <sup>2</sup>Virion Therapeutics, LLC, Newark, DE, USA

### INTRODUCTION

- Despite HBV preventative vaccines, chronic HBV infection remains a high unmet need:
- Most of the ~296M people with chronic HBV go undiagnosed and untreated<sup>1</sup>
- 1 in 4 people with chronic HBV will die prematurely from liver cirrhosis, HCC, or liver failure<sup>2</sup>
- Antivirals rarely achieve functional cure and require lifelong therapy<sup>3,4</sup>
- Chronic HBV infection is characterized by highly impaired and exhausted CD8+ T cells
- Current investigational treatments target viral and antigen suppression or immune modulation in the hope of restoring the same immune response that failed previously
- Restoring the same failed CD8+ T cell response is limited by:
- Irreversible epigenetic changes and terminal exhaustion of CD8+ T cells. which can blunt antiviral, small molecule, and checkpoint inhibitor interventions Vaccines aimed at:
- stimulating memory CD8+ T cells are unable to expand those that are already exhauste - naïve HBV-specific CD8+ T cells have a limited number of available
- naïve cells, since most people with a prolonged infection have already had these cells activated
- AdC6-gDHBV2 (VRON-0200) is a therapeutic vaccine for HBV functional cure that contains a genetically encoded checkpoint modifier, glycoprotein D (gD),<sup>5–8</sup> designed to enhance, broaden, and prolong CD8+ T cell responses to HBV core and polymerase
- gD lowers the activation threshold by inhibiting binding of the B and T lymphocyte attenuator (BTLA) to the herpes virus entry mediator (HVEM) on APCs (Fig 1)
- This results in T cell activation to sub-dominant epitopes<sup>9</sup>
- These T cells are generally not stimulated by an infection,<sup>10,11</sup> are less susceptible to suppression or exhaustion, and thereby remain functional and work for longer periods of time<sup>12</sup>
- We hypothesized that by eliminating HBV-infected hepatocytes, S antigen declines would occur without directly targeting S by vaccine-induced T cells

### AIM

To evaluate the immunogenicity and correlations between intrahepatic CD8+ T cell responses and HBV viral load and S antigen declines in blood in an AAV mouse model following a single i.m. injection of AdC6-gDHBV2



## **METHODS: IMMUNOGENICITY**

- A chimpanzee adenovirus vector of serotype 6 (AdC6) was used to express sequences of:
- Polymerase and core within gD (AdC6-gDHBV2)
- Polymerase and core without gD (AdC6-HBV2)
- Controls no insert, HIV gag (without gD), or unvaccinated animals
- C57BI/6 mice (groups of 5–10 animals) received a single i.m. dose of 1 x 10<sup>10</sup> vp of AdC6-gDHBV2, Ad6C-HBV2, or a control treatment
- The frequencies of insert-specific CD8+ T cells were determined in blood and spleen by intracellular cytokine staining (ICS) for IFN<sub>Y</sub> 4 and 8 weeks after injection, respectively

#### RESULTS

#### **IMMUNOGENICITY: NON-AAV-TREATED ANIMALS**

#### CD8+ T cell responses AdC6-gDHBV2 AdC6-HBV2 AdC6-gag Pie slice represents a T cell response to an individual peptide; the size of the slice is relative to the frequency 67% of response. 51/76 eptides Responses to individua peptides gated at 0.1% \*p-values between 0.001–0.01; \*\*\*p-values between 0.0001–0.001; \*\*\*\*p-values <0.0001 via 2-way ANOVA. AAV, adeno-associated virus; AdC6-qDHBV2 = VRON-0200; ANOVA, analysis of variance; IFNy, interferon-gamma

### **IMMUNOGENICITY: AAV-TREATED ANIMALS**

- animals (low-dose AAV8-1.3HBV study) (**Fig 3A**)
- groups (data not shown)



- Breadth and specificity of CD8+ T cell responses to individual peptides within a target sequence were performed via epitope mapping of splenocytes (CD8+ T cells tested by ICS for IFNγ) 8 weeks after treatment
- Immunogenicity was tested in the presence and absence of an AAV8-1.3HBV challenge (described below)
- In the AAV8-1.3HBV challenge studies:
- T cells from livers and spleens were tested at Week 12 post-vaccination • Frequencies and phenotype of CD8+ T cells to one immunodominant epitope within the N-terminus of Pol were tested for by staining with an MHC I tetramer and antibodies to various markers

• AdC6-gDHBV2-vaccinated animals had significantly higher frequencies of vaccine-induced IFNγ-producing CD8+ T cells in blood (Fig 2A) and were >2x as frequent, recognizing a higher number of epitopes in spleens (Fig 2B/C) than those observed in AdC6-HBV2-treated mice • Responses to some individual peptides were comparable between treatments, while others were only elicited by the AdC6-gDHBV2 vaccine (Fig 2D) • No T cell responses were observed in splenocytes of AdC6-HIV gag control-vaccinated animals (data not shown)



• Similar frequencies of vaccine-induced IFNγ-producing CD8+ T cells were observed in the blood of AdC6-gDHBV2- and AdC6-HBV2-vaccinated

• Significantly higher vaccine-induced IFNγ-producing CD8+ T cells were observed in the livers and spleens of AdC6-gDHBV2-vaccinated animals by both ICS and tetramer stain (**Fig 3B/C**) (high-dose AAV8-1.3HBV-treated groups)

• CD4+ T cell responses in vaccinated animals in both AAV8-1.3HBV and non-AAV8-1.3HBV studies were minimal or non-detectable in all treatment



### **METHODS: EFFICACY**

#### AAV8-1.3HBV viral dynamics study

- C57Bl/6 mice (n=5–10) were challenged intravenously via their tail vein with 1 x 10<sup>9</sup> (low-dose) or 1 x 10<sup>10</sup> (high-dose) viral genomes (vg) of AAV8-1.3HBV and 4 weeks later received a single i.m. injection of 1 x 10<sup>10</sup> vp of AdC6-gDHBV2, AdC6-HBV2, or no vaccination
- HBV DNA viral load and HBsAg titers (high-dose AAV8-1.3HBV study only) were evaluated in sera at different time points out to 16 weeks postvaccination by qPCR and ELISA, respectively; pre- and post-vaccination serum HBV DNA (copies/mL) and HBsAg levels (IU/mL) are reported

#### **Correlations between intrahepatic lymphocytes and splenocytes and viral dynamics**

• The relationship between HBV DNA viral load and HBsAg titers in sera at Week 16 post vaccination and the frequencies of vaccine-induced IFNγ-producing CD8+ T cells in liver and spleens were assessed by Spearman's rank correlations in high-dose AAV8-1.3HBV-treated animals

#### **ANTIVIRAL ACTIVITY**

- Following a single i.m. injection:
- AdC6-qDHBV2: HBV viral load declines of >3 log10 copies/mL were observed at Week 8/12 post-vaccination in both the low- (n=10) and high-dose (n=5) AAV8-1.3HBV mice upon vaccination (Fig 4A)
- AdC6-HBV2 and unvaccinated (control) mice: In contrast, these mice experienced <1.5 log10 copies/mL, and no change or increase in viral loads, respectively (**Fig 4A**)
- Levels of HBsAg showed:
- Increases in unvaccinated (control) mice
- Significant decreases by Week 12 after vaccination in both the AdC6-gDHBV2- and AdC6-HBV2-treated groups (**Fig 4B**)
- HBsAg antibodies at Week 16:
- Detected in all (100%) of AdC6-gDHBV2-vaccinated mice (n=5), albeit at low levels
- In contrast, only 1 of 5 (20%) of AdC6-HBV2-vaccinated animals had a detectable level (data not shown)

#### Figure 4. Viral dynamics

#### AAV8-1.3HBV challenge



AV, adeno-associated virus; AdC6-gDHBV2 = VRON-0200; ANOVA, analysis of variance; BL, baseline, HBsAg, hepatitis B surface antigen; vg, viral genomes.

#### **CORRELATIONS BETWEEN INTRAHEPATIC AND** SPLENIC LYMPHOCYTES AND VIRAL DYNAMICS

- At Week 16, the frequencies of vaccine-induced HBV-specific CD8+ T cells in spleens or livers strongly inversely correlated with HBV DNA viral load (Fig 5A)
- Other correlations are shown in **Fig 5B** • HBsAg levels inversely correlated with CD8+ T cell frequencies in both the liver and spleen and directly correlated with HBV DNA copies/mL in the blood

#### Figure 5. Correlations between Intrahepatic ymphocytes and splenocytes and viral dynamics

#### **Correlations between viral loads and CD8+ T cell** frequencies (high-dose AAV)



IFNy, interferon gamma; tet, tetramer; vg, viral genomes; Wk, week.

## Viral Hepatitis: **Experimental and** Pathophysiology Abstract 1285; TOP-107



EASL Congress Meeting Vienna, Austria 21–24 June 2023

**Splenocytes** R<sup>2</sup> = -0.81 Rank ICS IFNγ+CD8+ spleen IFNy+CD8+ liver Tet+CD8+ liver Antibodies 0.0471 0.0080 0.0116 0.6454 0.6482 0.7555 0.3892 0.1964 0.0149 0.0234 0.0692 0.0222 0.0238 0.0000 0.0002 0.1699 0.2425 0.2838 0.1917 0.0155 0.0003 0.0002 0.0070 0.0027 0.0196 Viral load, Wk 16 – 0.4359 0.5235 0.7212 0.0759 0.0057 0.0000 0.0000 0.0001 0.0000 0.0044 HBsAg, baseline – 0.0000 0.0000 0.1021 0.4918 0.3148 0.5424 0.5555 0.3796 0.8440 HBsAg, Wk 8 – 0.0000 0.0888 0.3469 0.3724 0.7020 0.6410 0.4190 0.7752 HBsAg, Wk 11 – 0.0379 0.2380 0.5500 0.9558 0.9558 0.7752 0.4357 HBsAg, Wk 14 – 0.0001 0.1198 0.0288 0.1082 0.0944 0.0527 HBsAg, Wk 16 – 0.0106 0.0019 0.0156 0.0116 0.0565

IFNγ+CD8+ liver – 0.0001 0.0000 0.0007

Tet+CD8+ spleen – 0.0000 0.0005

Tet+CD8+ liver – 0.0020

Antibodies

### CONCLUSIONS

- These are the first preclinical HBV therapeutic vaccine data that show correlations between intrahepatic CD8+ T cells and HBV viral load declines in serum
- Clearance of infected hepatocytes with AdC6-gDHBV2 correlates with Iower HBsAg levels
- The addition of gD, a novel checkpoint modifier of early T cell activation:
- Markedly enhanced the breadth of CD8+ T cell responses
- Significantly improved T cell frequencies within the liver
- A Phase 1b clinical trial of AdC6-gDHBV2 for HBV functional cure is scheduled to begin in Q3 2023

### REFERENCES

- WHO Hepatitis B Report; June 24, 2022
- Lok ASF, McMahon BJ. Hepatology 2009;50:661-2 Tsounis EP, et al. World J Gastroenterol 2021;27:2727–57
- Wong GLH, et al. J Hepatol 2022;76:1249–62.
- Luber A. et al. ESMO TAT 2021:Abstract 143.
- Xiang ZQ, et al. ASCO-SITC Clinical Immuno-Oncology Symposium 2020: Abstract 71.
- Stiles KM. et al. *J Virol* 2010:84:11646–60.
- Virion Therapeutics. Data on file. Zhang Y, Ertl HC. J Immunol 2014:193:1836–46.
- 10. Yewdell JW. Bennink JR. Annu Rev Immunol 1999;17:51–88.
- 11. Kotturi MF, et al. J Immunol 2008;181:2124–33
- 12. Rytelewski M, et al. PLoS One 2014;9:e90439.

### **ABBREVIATIONS**

AAV, adeno-associated virus; Ad, adenovirus; AdC6, chimpanzee adenoviral vector; ANOVA, analysis of variance; APC, antigen-presenting cell; BL, baseline; BTLA, B and T lymphocyte attenuator; CD, cluster of differentiatio ELISA, enzyme-linked immunosorbent assay; gD, glycoprotein D; HBsAg, hepatitis B surface antigen; HBV, hepatitis B virus; HCC, hepatocellular carcinoma; HIV gag, human immunodeficiency virus group specific antigen; HVEM, herpes virus entry mediator; ICS, intracellular cytokine staining; IFNy, interferon-gamma; i.m., intramuscular; LIGHT, lymphotoxin-like, exhibits inducible expression, and competes with herpes simplex virus glycoprotein D for HVEM, a receptor expressed by T lymphocytes; MHC, major histocompatibility complex; pol, polymerase; qPCR, quantitative polymerase chain reaction; SEM, standard error of the mean; TCR, T cell receptor; tet, tetramer; vg, viral genomes; vp, viral particles; VRON-0200 is AdC6-gDHBV2.

### ACKNOWLEDGEMENTS

We would like to thank The Ertl Laboratory at The Wistar Institute. All authors contributed to, and approved, the poster Editorial assistance was provided by Alison Lovibond, PhD, of BOLDSCIENCE Inc. and was funded by Virion Therapeutics, LLC. Funding for this study was provided by Virion Therapeutics, LLC.



### DISCLOSURES

Dr A Luber is CEO, Virion Therapeutics, LLC and owns shares in the company. He has no other conflicts to report.

#### FOR MORE INFORMATION

Contact Dr A Luber at: aluber@viriontx.com & www.VirionTx.com for permission to reprint and/or distribute.